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(FILE 'HOME' ENTERED AT 13:27:09 ON 09 MAY 2001)

L1 FILE 'REGISTRY' ENTERED AT 13:27:19 ON 09 MAY 2001  
1 S 61116-22-1/RN

FILE 'CAPLUS' ENTERED AT 13:28:29 ON 09 MAY 2001

L2 FILE 'REGISTRY' ENTERED AT 13:28:36 ON 09 MAY 2001  
SET SMARTSELECT ON  
SEL L1 1- CHEM : 8 TERMS  
SET SMARTSELECT OFF

L3 FILE 'CAPLUS' ENTERED AT 13:28:37 ON 09 MAY 2001  
1016 S L2

E DICARBOXYLIC ACID/CT  
E DICARBOXYLIC ACIDS/CT  
E E3 + ALL  
E MONOCARBOXYLIC ACIDS/CT  
E E3 + ALL  
E CARBOXYLIC ACIDS/CT

L4 425 S CANDIDA MALTOSA OR CANDIDA CLOACAE OR CANDIDA NOVELLUS OR

CAN

L5 7 S L3 (L) L4

L6 7 DUP REM L5 (0 DUPLICATES REMOVED)

=> s 61116-22-1/rn  
L1 1 61116-22-1/RN

=> d

L1 ANSWER 1 OF 1 REGISTRY COPYRIGHT 2001 ACS  
RN 61116-22-1 REGISTRY  
CN Oxidase, acyl coenzyme A (9CI) (CA INDEX NAME)  
OTHER NAMES:  
CN Acyl coenzyme A oxidase  
CN Acyl-CoA oxidase  
CN E.C. 1.3.3.6  
CN Fatty acyl-CoA oxidase  
CN Fatty acyl-coenzyme A oxidase  
CN Long-chain acyl-CoA oxidase  
CN Medium-chain acyl-CoA oxidase  
MF Unspecified  
CI MAN  
LC STN Files: AGRICOLA, ANABSTR, BIOBUSINESS, BIOSIS, BIOTECHNO, CA,  
CAPLUS, CHEMCATS, CSCHEM, EMBASE, MSDS-OHS, PROMT, TOXLIT, USPATFULL

\*\*\* STRUCTURE DIAGRAM IS NOT AVAILABLE \*\*\*

772 REFERENCES IN FILE CA (1967 TO DATE)

1 REFERENCES TO NON-SPECIFIC DERIVATIVES IN FILE CA

774 REFERENCES IN FILE CAPLUS (1967 TO DATE)

=> d ibib ab hit 1-7

L6 ANSWER 1 OF 7 CAPLUS COPYRIGHT 2001 ACS

ACCESSION NUMBER: 1998:6010 CAPLUS

DOCUMENT NUMBER: 128:125658

TITLE: Effect of cultivation conditions on the level of enzymes of n-alkane metabolism in *Candida maltosa* cells

AUTHOR(S): Sharyshev, A. A.; Peskova, E. V.; Komarova, G. N.

CORPORATE SOURCE: Institute of Biochemistry and Physiology of Microorganisms, Russian Academy of Sciences, Pushchino, 142292, Russia

SOURCE: Microbiology (Moscow) (Transl. of Mikrobiologiya) (1997), 66(6), 652-656

CODEN: MIBLAO; ISSN: 0026-2617

PUBLISHER: MAIK Nauka/Interperiodica Publishing

DOCUMENT TYPE: Journal

LANGUAGE: English

AB The effect of cultivation conditions on the content of cytochromes and the

activities of the key enzymes of n-alkane metab. was studied in the yeast *Candida maltosa*. Stationary-phase cells, grown on various substrates, were found to exhibit similar levels of cytochromes and enzymes.

IT 544-76-3, Hexadecane 9001-16-5 9007-43-6, Cytochrome c, biological studies 9035-37-4, Cytochrome b 9035-51-2, Cytochrome P 450, biological studies 9045-78-7, Isocitrate lyase 9073-63-6, Alcohol oxidase 61116-22-1, **Acyl-CoA oxidase**

RL: BSU (Biological study, unclassified); BIOL (Biological study) (effect of cultivation conditions on level of enzymes of n-alkane metab. in *Candida maltosa* cells)

L6 ANSWER 2 OF 7 CAPLUS COPYRIGHT 2001 ACS

ACCESSION NUMBER: 1995:706945 CAPLUS

DOCUMENT NUMBER: 123:106044

TITLE: Fatty acid oxidation enzymes of the yeast *Candida cloacae*

AUTHOR(S): West, Mark A.; Hill, Judy; Watson, Martin; Simon, William; Lindner, Nigel; Casey, John; Slabas, Antoni R.

CORPORATE SOURCE: Department Biological Sciences, University Durham, Durham, DH1 3LE, UK

SOURCE: Plant Lipid Metab., [Pap. Int. Meet. Plant Lipids], 11th (1995), Meeting Date 1994, 268-70. Editor(s): Kader, Jean-Claude; Mazliak, Paul. Kluwer:

Dordrecht,

Neth.

CODEN: 61OZAO

DOCUMENT TYPE: Conference

LANGUAGE: English

AB Cultures of parent and mutant strains of *C. cloacae* were grown under lab. conditions, and their induction patterns were compared over time for several enzymes of fatty acid oxidn. Alc. oxidase, an enzyme in the .omega.-oxidn. pathway, was purified 35-fold. The protein had a Mr of 70,000 and oxidized long-chain fatty alcs.

IT 9074-19-5, Hydratase 61116-22-1, **Acyl-CoA oxidase**

RL: BPR (Biological process); BIOL (Biological study); PROC (Process) (induction and purifn. of fatty acid oxidn. enzymes of *Candida cloacae*)

L6 ANSWER 3 OF 7 CAPLUS COPYRIGHT 2001 ACS

ACCESSION NUMBER: 1996:58714 CAPLUS  
DOCUMENT NUMBER: 124:166891  
TITLE: Cloning and characterization of the POX2 gene in  
Candida maltosa  
AUTHOR(S): Masuda, Yutaka; Park, Sun Mee; Ohta, Akinori; Takagi,  
Masamichi  
CORPORATE SOURCE: Tokyo, 113, Japan  
SOURCE: Gene (1995), 167(1/2), 157-61  
CODEN: GENED6; ISSN: 0378-1119  
DOCUMENT TYPE: Journal  
LANGUAGE: English

AB To study the function of **acyl-CoA oxidase** in  
an n-alkane-assimilating yeast, **Candida maltosa**, the  
authors isolated the POX2 gene which is a member of the **acyl-  
CoA oxidase** gene family. POX2 had a 2172-bp open  
reading frame (ORF) encoding an approx. 84-kDa polypeptide (724 amino  
acids (aa)) and was contiguous to POX4, another member of the **acyl-  
CoA oxidase** gene family on the same chromosomal DNA  
in a convergent arrangement. Northern blot anal. revealed that the  
expression of POX2 was induced in cells grown on oleic acid,  
n-tetradecanol and n-tetradecane. By using a gene-disruption technique,  
the authors constructed strains (termed P2DD and P4DD) in which both  
alleles of POX2 and POX4 were disrupted. The P2DD strain was normal in  
assimilation of various hydrophobic carbon sources, such as

n-tetradecane,

n-tetradecanol and oleic acid. In contrast, the P4DD strain was  
defective

in its ability to grow on such hydrophobic carbon sources.

AB To study the function of **acyl-CoA oxidase** in  
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n-tetradecane,

n-tetradecanol and oleic acid. In contrast, the P4DD strain was  
defective

in its ability to grow on such hydrophobic carbon sources.

ST sequence gene POX2 **Candida maltosa**; **acyl  
CoA oxidase** gene sequence Candida

IT 61116-22-1, Oxidase, acyl coenzyme A

RL: BSU (Biological study, unclassified); PRP (Properties); BIOL  
(Biological study)

(cloning, characterization and sequence of the POX2 gene in  
**Candida maltosa**)

L6 ANSWER 4 OF 7 CAPLUS COPYRIGHT 2001 ACS

ACCESSION NUMBER: 1989:167156 CAPLUS  
DOCUMENT NUMBER: 110:167156  
TITLE: Expression and transport of **Candida tropicalis**  
peroxisomal **acyl-coenzyme**

**A oxidase in the yeast  
Candida maltosa**

AUTHOR(S): Kamiryo, Tatsuyuki; Sakasegawa, Yuji; Tan, Hironobu  
 CORPORATE SOURCE: Fac. Integr. Arts Sci., Hiroshima Univ., Hiroshima,  
 730, Japan  
 SOURCE: Agric. Biol. Chem. (1989), 53(1), 179-86  
 CODEN: ABCHA6; ISSN: 0002-1369  
 DOCUMENT TYPE: Journal  
 LANGUAGE: English

AB The genes POX2 and POX4, which encode the subunits (PXP-2 and PXP-4) of peroxisomal fatty acyl-CoA oxidase of *C. tropicalis*, were introduced into the related yeast *C. maltosa*. The cells transformed with POX2 or POX4 showed high level expression of PXP-2 or PXP-4 in the purified peroxisomes. The polypeptides assocd. with the heterologous organelle were resistant to added protease, implying that they were transported into the peroxisomes. Truncated genes for PXP-4 were constructed in vitro and introduced into the host cells. Peptide-C, the COOH-terminal two-thirds of PXP-4, was efficiently transported into the host peroxisomes, and the polypeptide contg. the NH2-terminal one-third was also, in much lesser amt. These and other results suggested that there were at least 2 regions of peroxisomal targeting information in PXP-4 and the primary information was internal. Deletions in Peptide-C inhibited the transported of many, but not all, of the host-cell peroxisomal polypeptides. This suggested the existence of heterogeneous transport systems on the peroxisomal membrane.

TI Expression and transport of *Candida tropicalis* peroxisomal **acyl-coenzyme A oxidase** in the yeast  
**Candida maltosa**

IT **Candida maltosa**  
 (expression and transport of *Candida tropicalis* **fatty acyl-CoA oxidase** subunits in)

IT *Candida tropicalis*  
 (**fatty acyl-CoA oxidase** of peroxisome of, expression and transport in **Candida maltosa** of subunits of)

IT Peroxisome  
 (**fatty acyl-CoA oxidase** subunits of, of *Candida tropicalis*, expression and transport in **Candida maltosa** of)

IT Biological transport  
 (of **fatty acyl-CoA oxidase** of *Candida tropicalis*, into **Candida maltosa** peroxisomes, peroxisomal targeting sequences in relation to)

IT Molecular cloning  
 (of *Candida tropicalis* peroxisomal **fatty acyl-CoA oxidase** subunit genes, in **Candida maltosa**, peroxisomal transport in relation to)

IT Gene and Genetic element, microbial  
 RL: BIOL (Biological study)  
 (POX2, for **fatty acyl-CoA oxidase** subunit, of *Candida tropicalis* peroxisome, expression in **Candida maltosa** of)

IT Gene and Genetic element, microbial  
 RL: BIOL (Biological study)  
 (POX4, for **fatty acyl-CoA oxidase** subunit, of *Candida tropicalis* peroxisome, expression in **Candida maltosa** of)

IT 61116-22-1, **Fatty acyl-coenzyme**

**A oxidase**

RL: PRP (Properties)

(of *Candida tropicalis* peroxisome, expression and transport in  
*Candida maltosa* of subunits of)

L6 ANSWER 5 OF 7 CAPLUS COPYRIGHT 2001 ACS

ACCESSION NUMBER: 1988:449343 CAPLUS

DOCUMENT NUMBER: 109:49343

TITLE: Complete nucleotide sequence of the peroxisomal  
**acyl CoA oxidase** from the  
alkane-utilizing yeast **Candida**  
**maltosa**

AUTHOR(S): Hill, David E.; Boulay, Richard; Rogers, David

CORPORATE SOURCE: Genet. Inst., Cambridge, MA, 02140, USA

SOURCE: Nucleic Acids Res. (1988), 16(1), 365-6

CODEN: NARHAD; ISSN: 0305-1048

DOCUMENT TYPE: Journal

LANGUAGE: English

AB A 2127-nucleotide open reading frame of *C. maltosa* which codes for a  
709-amino acid acyl CoA oxidase (AOX) was sequenced. The gene is 81%  
similar to the AOX gene of *C. tropicalis*.

TI Complete nucleotide sequence of the peroxisomal **acyl CoA**  
**oxidase** from the alkane-utilizing yeast **Candida**  
**maltosa**

IT **Candida maltosa**

(**acyl CoA oxidase** gene of, nucleotide and  
encoded peptide sequences of)

IT Peroxisome

(**acyl CoA oxidase** of, of **Candida**

**maltosa**, gene for, nucleotide and encoded peptide sequences)

IT Gene and Genetic element, microbial

RL: BIOL (Biological study)

(for **acyl CoA oxidase**, of **Candida**

**maltosa**, nucleotide and encoded peptide sequences of)

IT Protein sequences

(of **acyl CoA oxidase**, of **Candida**

**maltosa**, complete)

IT Deoxyribonucleic acid sequences

(**acyl CoA oxidase**-specifying, of

**Candida maltosa**, complete)

IT 61116-22-1, **Acyl CoA oxidase**

RL: PRP (Properties)

(gene for, of **Candida maltosa**, nucleotide and  
encoded peptide sequences of)

L6 ANSWER 6 OF 7 CAPLUS COPYRIGHT 2001 ACS

ACCESSION NUMBER: 1981:170259 CAPLUS

DOCUMENT NUMBER: 94:170259

TITLE: Purification of acylcoenzyme A oxidase

PATENT ASSIGNEE(S): Amano Pharmaceutical Co., Ltd., Japan

SOURCE: Jpn. Kokai Tokkyo Koho, 8 pp.

CODEN: JKXXAF

DOCUMENT TYPE: Patent

LANGUAGE: Japanese

FAMILY ACC. NUM. COUNT: 1

PATENT INFORMATION:

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
JP 56008684	A2	19810129	JP 1979-81324	19790629

JP 60013664 B4 19850409

AB **Acyl-CoA oxidase** (I) is purified by adsorption on a water-insol. carrier contg.  $\text{CH}_3(\text{CH}_2)_n$  or  $\text{NH}_2(\text{CH}_2)_m$  groups, where  $n = 1-3$  and  $m = 2-8$ . Thus, an enzyme soln. prepd. from a culture of **Candida maltosa** IAM 12247 was dialyzed against 50 mM phosphate buffer (pH 8.0) and charged to a column of  $\text{CH}_3(\text{CH}_2)_3$ -Sephadex buffered to pH 8.0 with the phosphate buffer. The column was washed with the phosphate buffer contg. 0.1M NaCl and enzyme was eluted with the buffer contg. 0.5M NaCl. The eluate was mixed with  $(\text{NH}_4)_2\text{SO}_4$  at 80% satn. and the resulting ppt. was dialyzed and lyophilized to yield purified I (11.3 units/mg) free from catalase, other oxidases, lipase, and esterase.

AB **Acyl-CoA oxidase** (I) is purified by adsorption on a water-insol. carrier contg.  $\text{CH}_3(\text{CH}_2)_n$  or  $\text{NH}_2(\text{CH}_2)_m$  groups, where  $n = 1-3$  and  $m = 2-8$ . Thus, an enzyme soln. prepd. from a culture of **Candida maltosa** IAM 12247 was dialyzed against 50 mM phosphate buffer (pH 8.0) and charged to a column of  $\text{CH}_3(\text{CH}_2)_3$ -Sephadex buffered to pH 8.0 with the phosphate buffer. The column was washed with the phosphate buffer contg. 0.1M NaCl and enzyme was eluted with the buffer contg. 0.5M NaCl. The eluate was mixed with  $(\text{NH}_4)_2\text{SO}_4$  at 80% satn. and the resulting ppt. was dialyzed and lyophilized to yield purified I (11.3 units/mg) free from catalase, other oxidases, lipase, and esterase.

IT **Candida maltosa**  
(acyl CoA oxidase of)

IT **61116-22-1P**  
RL: PREP (Preparation)  
(of **Candida maltosa**, purifn. and properties of)

L6 ANSWER 7 OF 7 CAPLUS COPYRIGHT 2001 ACS

ACCESSION NUMBER: 1981:190327 CAPLUS

DOCUMENT NUMBER: 94:190327

TITLE: Acyl coenzyme A oxidase

PATENT ASSIGNEE(S): Amano Pharmaceutical Co., Ltd., Japan

SOURCE: Jpn. Kokai Tokkyo Koho, 6 pp.  
CODEN: JKXXAF

DOCUMENT TYPE: Patent

LANGUAGE: Japanese

FAMILY ACC. NUM. COUNT: 1

PATENT INFORMATION:

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
JP 56008683	A2	19810129	JP 1979-81323	19790629

AB **Acyl CoA oxidase** (I) [61116-22-1] is produced by culturing yeast (esp. **Candida**), fungi, or *Streptomyces*. Thus, *C. maltosa* IAM 12247 was cultured with shaking at 30.degree. for 24 h on a medium (pH 5.2) contg. n-alkane mixt. 1,  $(\text{NH}_4)_2\text{H}_2\text{PO}_4$  0.5,  $\text{KH}_2\text{PO}_4$  0.25,  $\text{MgSO}_4$  0.1, and yeast ext. 0.25%. The cells (22 g) were suspended in 50 mM phosphate buffer (pH 7.2) and disintegrated with a Dyno mill. The homogenate was mixed with  $(\text{NH}_4)_2\text{SO}_4$  at 45% satn. and centrifuged. The resulting ppt. was extd. with 50 mM phosphate buffer (pH 7.2) contg. 0.5% Triton X-100. The ext. was subjected to column chromatog. on Sephadex G-25 and DEAE-cellulose to yield 150 mg yellow powd. I.

IT **Candida maltosa**

(acyl CoA oxidase prodn. with)  
IT Fermentation  
    (acyl CoA oxidase, with **Candida**  
        maltosa)  
IT 61116-22-1P  
    RL: BMF (Bioindustrial manufacture); BIOL (Biological study); PREP  
    (Preparation)  
        (manuf. of, with **Candida maltosa**)



L2 ANSWER 1 OF 1 REGISTRY COPYRIGHT 2001 ACS  
RN 9023-03-4 REGISTRY  
CN Reductase, cytochrome c (reduced nicotinamide adenine dinucleotide phosphate) (9CI) (CA INDEX NAME)  
OTHER NAMES:  
CN Cytochrome c reductase  
CN Cytochrome c reductase (NADPH)  
CN Cytochrome c reductase (NADPH-dependent)  
CN Cytochrome c reductase (reduced nicotinamide adenine dinucleotide phosphate)  
CN Dihydronicotinamide adenine dinucleotide phosphate-cytochrome c reductase  
CN E.C. 1.6.2.4  
CN FAD-cytochrome c reductase  
CN NADP-cytochrome c reductase  
CN NADPH-cytochrome c oxidoreductase  
CN NADPH-cytochrome c reductase  
CN NADPH-dependent cytochrome c reductase  
CN NADPH-ferricytochrome c oxidoreductase  
CN NADPH-ferrihemoprotein reductase  
CN Reduced nicotinamide adenine dinucleotide phosphate-cytochrome c reductase  
CN TPNH-cytochrome c reductase  
MF Unspecified  
CI MAN  
LC STN Files: AGRICOLA, BIOBUSINESS, BIOSIS, BIOTECHNO, CA, CAPLUS, EMBASE,  
PIRA, PROMT, TOXLINE, TOXLIT, ULIDAT, USPATFULL

\*\*\* STRUCTURE DIAGRAM IS NOT AVAILABLE \*\*\*

2887 REFERENCES IN FILE CA (1967 TO DATE)

3 REFERENCES TO NON-SPECIFIC DERIVATIVES IN FILE CA

2887 REFERENCES IN FILE CAPLUS (1967 TO DATE)

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## NiceZyme View of ENZYME: EC 1.6.2.4

Official Name	
NADPH--ferrihemoprotein reductase.	
Alternative Name(s)	
NADPH--cytochrome p450 reductase. TPNH(2) cytochrome c reductase. Ferrihemoprotein p450 reductase.	
Reaction catalysed	
$  \begin{array}{l}  \text{NADPH} \\  + \quad 2 \text{ ferricytochrome} \\  \rightleftharpoons \\  \text{NADP (+)} \\  + \quad 2 \text{ ferrocytochrome}  \end{array}  $	
Cofactor(s)	
FMN; FAD.	
Comments	
<ul style="list-style-type: none"> <li>Catalyses the reduction of heme-thiolate-dependent monooxygenases such as EC <u>1.14.14.1</u>, and is part of the microsomal hydroxylating system.</li> <li>Also reduces cytochrome b5 and cytochrome c.</li> </ul>	
Cross-References	
Biochemical Pathways; map number(s)	<u>T6</u> , <u>U6</u>
BRENDA	<u>1.6.2.4</u>
EMP/PUMA	<u>1.6.2.4</u>
WIT	<u>1.6.2.4</u>
KYOTO UNIVERSITY LIGAND CHEMICAL DATABASE	<u>1.6.2.4</u>
MEDLINE	Find literature relating to <u>1.6.2.4</u>
SWISS-PROT	<p>P14779, CPXB_BACME; P50126, NCPR_CANMA; P37201, NCPR_CANTR;  Q05001, NCPR_CATRO; P37039, NCPR_CAVPO; P16435, NCPR_HUMAN;  P37040, NCPR_MOUSE; Q07994, NCPR_MUSDO; P37116, NCPR_PHAU;  P04175, NCPR_PIC; P00380, NCPR_PABT; P00380, NCPR_PABT</p>

	<a href="#">P04173, NCPR_ETG;</a>	<a href="#">P00585, NCPR_KAB1;</a>	<a href="#">P00500, NCPR_KAT;</a>
	<a href="#">P19618, NCPR_SALTR;</a>	<a href="#">P36587, NCPR_SCHPO;</a>	<a href="#">P16603, NCPR_YEAST;</a>

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			<a href="#">China</a>	<a href="#">Korea</a>

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## BRENDA:1.6.2.4

E.C. number	1.6.2.4 (BRENDA <u>copyright</u> notice)
Original Organism	<p>#1# <u>Pig</u> &lt;1, 11, 14, 15, 29, 32, 35, 38, 47&gt;</p> <p>#2# <u>Rat</u> &lt;2, 5, 10, 16, 27, 29, 31, 33, 34, 36, 41, 43, 46&gt;</p> <p>#3# <u>Rabbit</u> &lt;3, 4, 30, 39&gt;</p> <p>#4# <u>Human</u> &lt;45&gt;</p> <p>#5# <u>Helianthus tuberosus</u> (L. var. Blanc commun., Jerusalem artichoke) &lt;6, 9&gt;</p> <p>#6# <u>Nitrobacter winogradskyi</u> &lt;7&gt;</p> <p>#7# <u>Hamster</u> &lt;8&gt;</p> <p>#8# <u>Trypanosoma cruzi</u> &lt;12&gt;</p> <p>#9# <u>Trichosporon cutaneum</u> &lt;13&gt;</p> <p>#10# <u>Lodderomyces elongisporus</u> &lt;17&gt;</p> <p>#11# <u>Aspergillus ochraceus</u> &lt;18&gt;</p> <p>#12# <u>Saccharomyces cerevisiae</u> (grown anaerobically) &lt;19, 23, 28, 37, 42&gt;</p> <p>#13# <u>Candida tropicalis</u> (grown on alkanes) &lt;20, 22, 26&gt;</p> <p>#14# <u>Spodoptera eridania</u> (southern Armyworm) &lt;21&gt;</p> <p>#15# <u>Catharanthus roseus</u> &lt;24&gt;</p> <p>#16# <u>Tetrahymena pyriformis</u> &lt;40&gt;</p> <p>#17# <u>Plants</u> (e.g. maize, potato, avocado, bramble, tulip, leek, Vicia faba, sunflower) &lt;6&gt;</p> <p>#18# <u>Housefly</u> &lt;44&gt;</p> <p>#19# <u>Horse</u> &lt;5&gt;</p> <p>#20# <u>Chicken</u> &lt;25&gt;</p>
Systematic name	NADPH:ferrihemoprotein oxidoreductase
Recommended name	NADPH-ferrihemoprotein reductase

<b>Synonyms</b>	<ul style="list-style-type: none"> <li>☞ Dihydroxynicotinamide adenine dinucleotide phosphate-cytochrome c reductase</li> <li>☞ EC 1.6.99.2 #2# (formerly) &lt;27&gt;</li> <li>☞ Aldehyde reductase (NADPH-dependent) #2# &lt;10&gt;</li> <li>☞ NADPH-cytochrome p-450 reductase #2# &lt;10, 27&gt;</li> <li>☞ FAD-cytochrome c reductase</li> <li>☞ NADP-cytochrome c reductase</li> <li>☞ NADPH-dependent cytochrome c reductase</li> <li>☞ NADPH-ferricytochrome c oxidoreductase</li> <li>☞ NADPH-cytochrome c oxidoreductase</li> <li>☞ Reduced nicotinamide adenine dinucleotide phosphate-cytochrome c reductase</li> <li>☞ TPNH-cytochrome c reductase</li> <li>☞ NADPH-cytochrome c reductase</li> <li>☞ Reductase, cytochrome c (reduced nicotinamide adenine dinucleotide phosphate)</li> <li>☞ Ferrihemprotein P450 reductase</li> <li>☞ TPNH2 cytochrome c reductase</li> <li>☞ NADP-cytochrome reductase</li> <li>☞ Cytochrome c reductase (reduced nicotinamide adenine dinucleotide phosphate, NADPH, NADPH-dependent)</li> </ul>
<b>CAS registration number</b>	9023-03-4
<b>Reaction</b>	NADPH + 2 ferricytochrome = NADP+ + 2 ferrocytochrome
<b>Reaction type</b>	Redox reaction

<b>Substrates/products</b>	<p>☑ S: NADPH + O<sub>2</sub> #<u>1,12</u># (slow reaction, presence of menadione, or duroquinone, or vitamin K3 essential) &lt;<u>14, 28</u>&gt; P: NADP<sup>+</sup> + O<sub>2</sub><sup>-</sup> #<u>1,12</u># (superoxide anion) &lt;<u>14, 28</u>&gt;</p> <p>☑ S: More #<u>1-3</u># (O-deethylation of 7-ethoxycoumarin, #<u>3</u># &lt;<u>30</u>&gt;; N-demethylation of benzphetamine, #<u>2,3</u># &lt;<u>30, 33</u>&gt;; aniline hydroxylase, #<u>2</u># &lt;<u>33</u>&gt;; as part of MEOS i.e. microsomal ethanol-oxidizing system composed of NADPH-cytochrome c reductase, Cytochrome P-450, phospholipids, #<u>2</u># &lt;<u>33</u>&gt;; omega-hydroxylation of fatty acids together with cytochrome P-450, #<u>1</u># &lt;<u>38</u>&gt;) &lt;<u>30, 33, 38</u>&gt; P: ?</p> <p>☑ S: 17-Hydroxyprogesterone + NADPH #<u>2</u># &lt;<u>34</u>&gt; P: Androstendione + 2-carbon fragment #<u>2</u># (removal of 2-carbon side chain from 17-position of 21-carbon steroids) &lt;<u>34</u>&gt;</p> <p>☑ S: NADPH + cinnamate #<u>5</u># &lt;<u>9</u>&gt; P: NADP<sup>+</sup> + p-coumarate #<u>5</u># &lt;<u>9</u>&gt;</p> <p>☑ S: NADPH + ferricytochrome c #<u>1-3,5-8,10,12-15,19</u># (NADH less than 5% of NADPH activity, #<u>1</u># &lt;<u>1</u>&gt;; additional electron acceptors: 2,6-dichlorophenolindophenol, #<u>1,2,5,6,8,10,12-15</u># &lt;<u>6, 7, 12, 14, 17, 20, 21, 24, 26, 28, 36</u>&gt;; cytochrome P-450, #<u>1,2,5,7,13</u># &lt;<u>1, 8, 9, 20, 36, 38</u>&gt;; ferricyanide, #<u>5,8,10,12-15</u># &lt;<u>6, 9, 12, 17, 20, 21, 24, 26, 28</u>&gt;; menadione, #<u>6,8,13</u># &lt;<u>7, 12, 20, 26</u>&gt;; neotetrazolium chloride, #<u>2,13</u># &lt;<u>20, 26, 36</u>&gt;; nitroblue tetrazolium salt, #<u>1</u># &lt;<u>14</u>&gt;; vitamin K3, #<u>2</u># &lt;<u>36</u>&gt;; benzoquinone, #<u>2</u># &lt;<u>36</u>&gt;) &lt;<u>1, 4-9, 12, 14, 17, 20, 21, 24, 26, 28, 36, 38</u>&gt; P: NADP<sup>+</sup> + ferrocyclochrome c</p> <p>☑ S: NADPH + hexadecanal #<u>2</u># (hexadecanol replaceable by p-nitroacetophenone, or p-pyridinecarboxaldehyde, or p-nitrobenzaldehyde) &lt;<u>10</u>&gt; P: NADP<sup>+</sup> + hexadecanol #<u>2</u># &lt;<u>10</u>&gt;</p>
<b>Natural substrates</b>	<p>☑ More #<u>2,3,13</u># (monooxygenase system composed of cytochrome P-450, NADPH-cytochrome c reductase, phospholipids, #<u>2,13</u># &lt;<u>16, 26</u>&gt;; detoxification of drugs, inactivation of procarcinogens, #<u>2</u># &lt;<u>16</u>&gt;; biotransformation of airborne compounds, #<u>3</u># &lt;<u>30</u>&gt;) &lt;<u>16, 26, 30</u>&gt;</p> <p>☑ NADPH + cytochrome P-450 #<u>5,17</u># &lt;<u>6</u>&gt;</p>
<b>Turnover number (1/min)</b>	<p>☑ 6100 #<u>13</u># {cytochrome c} &lt;<u>26</u>&gt;</p> <p>☑ 3870 #<u>8</u># {ferricyanide} &lt;<u>12</u>&gt;</p> <p>☑ 897 #<u>8</u># {cytochrome c} &lt;<u>12</u>&gt;</p> <p>☑ 458 #<u>8</u># {2,6-dichlorophenolindophenol} &lt;<u>12</u>&gt;</p> <p>☑ 87 #<u>8</u># {menadione} &lt;<u>12</u>&gt;</p> <p>☑ -999 #<u>1,2</u># &lt;<u>29, 31, 33</u>&gt;</p>

<b>Specific activity (micromol/min/mg)</b>	<ul style="list-style-type: none"> <li>150-180 #12# &lt;28&gt;</li> <li>63.8 #2# &lt;16&gt;</li> <li>40 #19# &lt;5&gt;</li> <li>15.2 #1# &lt;1&gt;</li> <li>-999 #1-3,5-7,9,13,15,20# &lt;3, 7, 8-11, 13, 14, 20, 24, 25, 27, 30, 34, 36&gt;</li> </ul>
<b>Km-value (mM)</b>	<ul style="list-style-type: none"> <li>7.2 #2# {ethanol} (microsomal ethanol oxidizing system) &lt;33&gt;</li> <li>2.5 #2# {benzalacetone} &lt;10&gt;</li> <li>1.4 #2# {p-nitroacetophenone} &lt;10&gt;</li> <li>0.31 #2# {p-nitrobenzaldehyde} &lt;10&gt;</li> <li>0.077 #13# {2,6-dichlorophenolindophenol} &lt;26&gt;</li> <li>0.03 #2# {hexadecanol} &lt;10&gt;</li> <li>0.022 #1# {NADPH} (similar values, #1,5,12,14# &lt;9, 14, 21, 28&gt;) &lt;11&gt;</li> <li>0.013 #2,14# {cytochrome c} (similar value, #1# &lt;32&gt;) &lt;5, 21&gt;</li> <li>0.0082 #19# {cytochrome c} &lt;5&gt;</li> <li>0.006 #1# {cytochrome c} (similar values, #5# &lt;9&gt;) &lt;35&gt;</li> <li>0.0053 #1# {menadione} &lt;14&gt;</li> <li>0.0036 #1# {NADPH} (similar values, #1,3,6,8,14,15,20# &lt;4, 7, 12, 21, 24, 25, 32, 35&gt;) &lt;1&gt;</li> <li>0.0019 #3# {azidonitrophenyl-gamma-aminobutyryl-NADPH} &lt;4&gt;</li> <li>-999 #1,8# (O2-generation, #1# &lt;14&gt;) &lt;11, 12, 14&gt;</li> </ul>
<b>pH-optimum</b>	<ul style="list-style-type: none"> <li>8-9 #1# &lt;35&gt;</li> <li>7.8 #13# &lt;20&gt;</li> <li>7.8-8 #12,20# &lt;25, 28&gt;</li> <li>7.7 #11# &lt;18&gt;</li> <li>7.5-9 #14# &lt;21&gt;</li> <li>7-7.4 #1# (O2- generation) &lt;14&gt;</li> <li>6.9-7.5 #2# (microsomal ethanol-oxidizing system) &lt;33&gt;</li> </ul>
<b>pH-range</b>	<ul style="list-style-type: none"> <li>7-8.5 #13# (less than 50% of maximal activity above and below) &lt;26&gt;</li> <li>6.5-9 #14# &lt;21&gt;</li> </ul>

<b>Cofactors/prosthetic groups</b>	<p>☒ FAD #1-3,5-8,10,12-15,20# (ratio FAD:FMN 1:1, #1,5,10,12-14# &lt;1, 9, 11, 17, 20, 21, 26, 28&gt;; 1 mol per subunit, #6,8,12# &lt;7, 12, 23&gt;; tightly bound, #12,13# &lt;20, 28&gt;; loosely bound, #12# &lt;42&gt;) &lt;1-3, 7-9, 11, 12, 17, 20, 21, 23-26, 28, 31-33, 35, 42&gt;</p> <p>☒ FMN #1-3,5,7,10,12-15,20# (ratio FAD:FMN 1:1, #1,5,10,12-14# &lt;1, 9, 11, 17, 20, 21, 26, 28&gt;; tightly bound, #13# &lt;20&gt;; loosely bound, #12# &lt;28, 42&gt;; 1 mol per mol of enzyme, #10,13# &lt;17, 20&gt;) &lt;1-3, 8, 9, 11, 17, 20, 21, 23-26, 28, 31, 32, 35, 42&gt;</p> <p>☒ Menadione #14# (slight stimulation) &lt;21&gt;</p> <p>☒ NADPH #1-3,5-8,10,12-15,17,19# (NADH less than 5% of NADPH activity, #1# &lt;1&gt;; not replaceable by NADH, #2# &lt;33&gt;) &lt;1, 4-9, 12, 14, 17, 20, 21, 24, 28, 33, 36, 38&gt;</p> <p>☒ Nonionic detergent #10# (activation) &lt;17&gt;</p>
<b>Metal ions/salts</b>	<p>More #5,15# (stimulation by increasing ionic strength) &lt;9, 24&gt;</p>
<b>Inhibitors</b>	<p>☒ 2'-AMP #14# &lt;21&gt;</p> <p>☒ 2,6-Dichlorophenol-indophenol #1# (formation of superoxide anion) &lt;14&gt;</p> <p>☒ 3-Aminonicotinamide adenine dinucleotide phosphate #5# &lt;6&gt;</p> <p>☒ 5,5'-Dithiobis(2-nitrobenzoate) #12# (in absence of FAD or NADPH) &lt;23&gt;</p> <p>☒ Alizarin #14# &lt;21&gt;</p> <p>☒ CO #2# &lt;33&gt;</p> <p>☒ HgCl<sub>2</sub> #12,13# &lt;20, 26, 28&gt;</p> <p>☒ High ionic strength #5# &lt;9&gt;</p> <p>☒ Mersalyl #3,14# &lt;3, 21&gt;</p> <p>☒ N<sub>2</sub> #2# &lt;33&gt;</p> <p>☒ NAD<sup>+</sup> #6# &lt;7&gt;</p> <p>☒ NADP<sup>+</sup> #1,5,6,13,14# &lt;6, 7, 9, 21, 26, 32&gt;</p> <p>☒ p-Chloromercuribenzoate #12-15# &lt;20, 21, 24, 26, 28&gt;</p> <p>☒ Sodium formate #2# &lt;33&gt;</p>



Source tissue	<ul style="list-style-type: none"> <li><input checked="" type="checkbox"/> Brain #2# &lt;10&gt;</li> <li><input checked="" type="checkbox"/> Bulbs #17# (tulip) &lt;6&gt;</li> <li><input checked="" type="checkbox"/> Cell #6# &lt;7&gt;</li> <li><input checked="" type="checkbox"/> Endometrium #1# &lt;15&gt;</li> <li><input checked="" type="checkbox"/> Epimastigotes #8# &lt;12&gt;</li> <li><input checked="" type="checkbox"/> Kidney #1,20# &lt;25, 32, 35&gt;</li> <li><input checked="" type="checkbox"/> Liver #1-3,7# &lt;2, 5, 8, 16, 27, 29, 31, 33, 34, 36, 38, 39, 43, 46, 47&gt;</li> <li><input checked="" type="checkbox"/> Lung #3# &lt;30&gt;</li> <li><input checked="" type="checkbox"/> Mesocarp #17# (avocado) &lt;6&gt;</li> <li><input checked="" type="checkbox"/> Midgut #14# (of larvae) &lt;21&gt;</li> <li><input checked="" type="checkbox"/> Peritoneal neutrophils #3# &lt;3&gt;</li> <li><input checked="" type="checkbox"/> Placenta #4,19# &lt;5, 45&gt;</li> <li><input checked="" type="checkbox"/> Polymorphonuclear leukocytes #1# &lt;1, 14&gt;</li> <li><input checked="" type="checkbox"/> Seedlings #15,17# (Vicia faba, leek, sunflower, #17# &lt;6&gt;) &lt;6, 24&gt;</li> <li><input checked="" type="checkbox"/> Spleen #1# &lt;35&gt;</li> <li><input checked="" type="checkbox"/> Testis #1# &lt;11&gt;</li> <li><input checked="" type="checkbox"/> Tuber #5# &lt;6, 9&gt;</li> </ul>
Localisation	<ul style="list-style-type: none"> <li><input checked="" type="checkbox"/> Cytosol #6,8# &lt;7, 12&gt;</li> <li><input checked="" type="checkbox"/> Endoplasmic reticulum #2# &lt;43&gt;</li> <li><input checked="" type="checkbox"/> membrane-bound #1,3,13,20# (outer membrane of mitochondria, #20# &lt;25&gt;) &lt;1, 3, 4, 14, 20, 25&gt;</li> <li><input checked="" type="checkbox"/> Microsomes #1,2,5,7,9-12,14,16,19# &lt;2, 5, 8-10, 13, 15-18, 21, 31, 33, 35-38, 40&gt;</li> <li><input checked="" type="checkbox"/> Nuclear envelope #2# &lt;27&gt;</li> <li><input checked="" type="checkbox"/> Nucleus #12# (low activity) &lt;19&gt;</li> <li><input checked="" type="checkbox"/> Spheroplasts #12# &lt;19&gt;</li> </ul>

<b>Purification</b>	<ul style="list-style-type: none"> <li>☑ #1# &lt;1, 11, 14, 29, 38, 47&gt;</li> <li>☑ #10# &lt;17&gt;</li> <li>☑ #12# &lt;23, 28&gt;</li> <li>☑ #13# &lt;20, 26&gt;</li> <li>☑ #14# &lt;21&gt;</li> <li>☑ #15# &lt;24&gt;</li> <li>☑ #19# &lt;5&gt;</li> <li>☑ #2# (FAD-depleted enzyme &lt;2&gt;) &lt;2, 5, 10, 16, 29, 33, 36, 41&gt;</li> <li>☑ #3# &lt;3, 30&gt;</li> <li>☑ #5# &lt;9&gt;</li> <li>☑ #6# &lt;7&gt;</li> <li>☑ #7# &lt;8&gt;</li> <li>☑ #8# &lt;12&gt;</li> </ul>
<b>Crystallisation</b>	<ul style="list-style-type: none"> <li>#12# (Saccharomyces cerevisiae) &lt;23&gt;</li> </ul>
<b>Molecular Weight</b>	<ul style="list-style-type: none"> <li>☑ 400000 #1# (pig, gel filtration) &lt;1&gt;</li> <li>☑ 100000 #8# (Trypanosoma cruzi, gel filtration) &lt;12&gt;</li> <li>☑ 82000-85000 #4,5,12,18# (Helianthus tuberosus, SDS-PAGE followed by Western blotting &lt;6&gt;; Saccharomyces cerevisiae, calculation from FAD content &lt;28&gt;; house fly &lt;44&gt;; human placenta &lt;45&gt;) &lt;6, 28, 44, 45&gt;</li> <li>☑ 78000-79000 #2,3# (rabbit, sedimentation equilibrium centrifugation &lt;39&gt;; rat liver &lt;46&gt;) &lt;39, 46&gt;</li> <li>☑ 70000 #1,5,12# (Nitrobacter winogradskyi, gel filtration &lt;7&gt;; pig testis, gel filtration &lt;11&gt;; Saccharomyces cerevisiae, gel filtration &lt;28&gt;) &lt;7, 11, 28&gt;</li> <li>☑ 65000-68000 #1,3,12,13# (Saccharomyces cerevisiae, sedimentation equilibrium centrifugation, values depending on pH &lt;23&gt;; Candida tropicalis, gel filtration, sedimentation equilibrium centrifugation &lt;26&gt;; pig kidney, gel filtration &lt;32, 35&gt;; rabbit liver, gel filtration &lt;39&gt;) &lt;23, 26, 32, 35, 39&gt;</li> <li>☑ -999 #5,13# (differences in MW partially due to method of solubilization) &lt;9, 20&gt;</li> </ul>

<b>Subunits</b>	<p>☐ ? #<u>1-3,5,7,10,13,14</u># (x * 72000-87000, pig, SDS-PAGE &lt;<u>1, 11, 14, 29</u>&gt;; rabbit, SDS-PAGE &lt;<u>3</u>&gt;; horse, SDS-PAGE &lt;<u>5</u>&gt;; rat, SDS-PAGE &lt;<u>5, 10, 16, 27, 29</u>&gt;; hamster, SDS-PAGE &lt;<u>8</u>&gt;; Helianthus tuberosus, SDS-PAGE &lt;<u>9</u>&gt;; Lodderomyces elongisporus, SDS-PAGE &lt;<u>17</u>&gt;; Candida tropicalis, SDS-PAGE &lt;<u>20</u>&gt;; Spodoptera eridania, SDS-PAGE &lt;<u>21</u>&gt;) &lt;<u>1, 3, 5, 8-11, 14, 16, 17, 20, 21, 27, 29</u>&gt;</p> <p>☐ Dimer #<u>6,8,12</u># (2 * 36000, Nitrobacter winogradskyi, SDS-PAGE &lt;<u>7</u>&gt;; 2 * 52000, Trypanosoma cruzi, SDS-PAGE &lt;<u>12</u>&gt;; 2 * 34300-40000, Saccharomyces cerevisiae, SDS-PAGE, sedimentation equilibrium centrifugation after treatment with guanidine-HCl &lt;<u>23</u>&gt;) &lt;<u>7, 12, 23</u>&gt;</p> <p>☐ Monomer #<u>3,12</u># (72000, Saccharomyces cerevisiae, SDS-PAGE &lt;<u>28</u>&gt;; 75000, rabbit, Triton-solubilized, SDS-PAGE &lt;<u>39</u>&gt;; 68000, rabbit, trypsin-solubilized, SDS-PAGE &lt;<u>39</u>&gt;) &lt;<u>28, 39</u>&gt;</p> <p>☐ More #<u>5,13</u># (differences in subunit weight partially due to method of solubilization) &lt;<u>9, 20</u>&gt;</p>
<b>Temperature stability (deg.C)</b>	<p>☐ 100 #<u>1</u># (10 min, inhibition of O<sub>2</sub>-formation) &lt;<u>14</u>&gt;</p> <p>☐ 60 #<u>2</u># (inactivation) &lt;<u>10</u>&gt;</p> <p>☐ 40 #<u>14</u># (50% activity) &lt;<u>21</u>&gt;</p> <p>☐ 36 #<u>14</u># (inactivation above) &lt;<u>21</u>&gt;</p> <p>☐ 25-30 #<u>2</u># (diluted solutions: gradual loss of activity) &lt;<u>41</u>&gt;</p> <p>☐ -999 #<u>12</u># (FAD and NADPH: protection against thermal inactivation) &lt;<u>23</u>&gt;</p>
<b>General stability</b>	<p>☐ #<u>12,13</u># FAD, FMN necessary for stabilization during purification &lt;<u>20, 28</u>&gt;</p> <p>☐ #<u>1</u># Instable during purification &lt;<u>32</u>&gt;</p>

<b>Storage stability</b>	<p>☞ #2# -15°C or -20°C, 10 mM phosphate buffer, pH 7.5, several months &lt;41&gt;</p> <p>☞ #14# -15°C, more than 1 year &lt;21&gt;</p> <p>☞ #3# -20°C, N<sub>2</sub>-atmosphere, several weeks &lt;30&gt;</p> <p>☞ #13# -70°C, 0.15 mM potassium phosphate buffer, pH 7, 1 mM mercaptoethanol, 1 mM EDTA, 1 micromol FMN, 1 micromol FAD, 0.3% Mülgofer BC-720, 30% glycerol, several months &lt;20&gt;</p> <p>☞ #2# -78°C, 30 mM potassium phosphate buffer, pH 7.7, 0.1 mM EDTA, 20% glycerol, 0.4 mM PMSF &lt;16&gt;</p> <p>☞ #2# -80°C &lt;10&gt;</p> <p>☞ #7# -80°C, 50 mM phosphate buffer, pH 7.4, 0.1 mM EDTA, 20% glycerol &lt;8&gt;</p> <p>☞ #8# -90°C or -20°C, 24 h, 5-10% loss of activity, reactivation by FAD &lt;12&gt;</p> <p>☞ #14# 0-4°C, several months &lt;21&gt;</p> <p>☞ #2# 0°C, some days &lt;41&gt;</p> <p>☞ #2# 4°C or room temperature, FAD-depleted enzyme &lt;2&gt;</p>
<b>Renaturated</b>	<p>#1,3,5# (reconstitution of O<sub>2</sub>- generating system, #3# &lt;3&gt;; reconstitution of monooxygenase system, #5# &lt;9&gt;) &lt;3, 9, 30, 38&gt;</p>
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BRENDA:1.6.2.4 Aspergillus ochraceus



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RN 9038-14-6 REGISTRY

CN Oxygenase, mono- (9CI) (CA INDEX NAME)

OTHER NAMES:

CN Baeyer-Villigerase

CN Cytochrome P 450 hydroperoxidase

CN Cytochrome P 450 monooxygenase

CN Cytochrome P 450-linked monooxygenase

CN Cytochrome P-450 mixed-function oxidase

CN E.C. 1.14.14.1

CN E.C. 1.14.14.2

CN Flavin monooxygenase

CN Flavin-containing monooxygenase

CN Flavin-contg. monooxygenase

CN Flavin-contg. monooxygenase 1

CN Flavin-contg. monooxygenase 3

CN Flavoprotein monooxygenase

CN Flavoprotein-linked monooxygenase

CN HCE hydroxylase

CN Microsomal monooxygenase

CN Mixed function monooxygenase

CN Mixed-function oxidase

CN Mixed-function oxygenase

CN Monooxygenase

CN Oxidase, mixed function

CN Oxygenase, flavoprotein-linked mono-

DR 9040-60-2, 55963-41-2, 62213-32-5

MF Unspecified

CI MAN

LC STN Files: AGRICOLA, ANABSTR, BIOBUSINESS, BIOSIS, BIOTECHNO, CA,  
CAPLUS, CASREACT, CEN, CIN, CSNB, EMBASE, IFICDB, IFIPAT, IFIUDB, PIRA,  
PROMT, TOXLINE, TOXLIT, ULIDAT, USPATFULL

\*\*\* STRUCTURE DIAGRAM IS NOT AVAILABLE \*\*\*

3903 REFERENCES IN FILE CA (1967 TO DATE)

22 REFERENCES TO NON-SPECIFIC DERIVATIVES IN FILE CA

3913 REFERENCES IN FILE CAPLUS (1967 TO DATE)

**BRENDA:1.14.14.1**

<b>E.C. number</b>	1.14.14.1 (BRENDA <a href="#">copyright notice</a> )
<b>Original Organism</b>	#1# <u>Mammals</u> (e.g. mouse <1, 2>; rabbit <3>. There is no clear evidence whether the enzymes isolated from microsomes of different species, or even from the same species by different research groups are identical) <1-3>
<b>Systematic name</b>	Substrate, reduced-flavoprotein:oxygen oxidoreductase (RH-hydroxylating or-epoxidizing)
<b>Recommended name</b>	Unspecific monooxygenase
<b>Synonyms</b>	<ul style="list-style-type: none"> <li><input checked="" type="checkbox"/> Microsomal monooxygenase</li> <li><input checked="" type="checkbox"/> EC 1.99.1.1 (formerly)</li> <li><input checked="" type="checkbox"/> EC 1.14.99.8 (formerly)</li> <li><input checked="" type="checkbox"/> EC 1.14.1.1 (formerly)</li> <li><input checked="" type="checkbox"/> Microsomal P-450</li> <li><input checked="" type="checkbox"/> Aryl hydrocarbon hydroxylase</li> <li><input checked="" type="checkbox"/> Aryl-4-monooxygenase</li> <li><input checked="" type="checkbox"/> Xenobiotic monooxygenase</li> <li><input checked="" type="checkbox"/> Flavoprotein-linked monooxygenase</li> <li><input checked="" type="checkbox"/> Flavoprotein monooxygenase</li> <li><input checked="" type="checkbox"/> Oxygenase, flavoprotein-linked mono-</li> <li><input checked="" type="checkbox"/> EC 1.14.14.2 (formerly)</li> </ul>
<b>CAS registration number</b>	62213-32-5
<b>Reaction</b>	RH + reduced flavoprotein + O <sub>2</sub> = ROH + oxidized flavoprotein + H <sub>2</sub> O
<b>Reaction type</b>	<ul style="list-style-type: none"> <li><input checked="" type="checkbox"/> Deamination</li> <li><input checked="" type="checkbox"/> Desulfation</li> <li><input checked="" type="checkbox"/> Epoxidation</li> <li><input checked="" type="checkbox"/> Hydroxylation</li> <li><input checked="" type="checkbox"/> N-, S-, O-Dealkylation</li> <li><input checked="" type="checkbox"/> N-Oxidation</li> <li><input checked="" type="checkbox"/> Redox reaction</li> <li><input checked="" type="checkbox"/> Reduction of azo, nitro, N-oxide groups</li> <li><input checked="" type="checkbox"/> Sulfoxidation</li> </ul>

<b>Substrates/products</b>	<p>☛ S: Aryl hydrocarbons + reduced flavoprotein + O<sub>2</sub> #1# (e.g. benzo[a]pyrene, ethoxyresufurin, biphenyl, p-nitroanisole, acetanilide, 2-acetylaminofluorene, 2-ethoxycoumarin, estradiol-17beta, testosterone, #1# &lt;1, 2&gt;; prostaglandins, #1# &lt;3&gt;) &lt;1-3&gt;</p> <p>P: ?</p> <p>☛ S: More #1# (acts on a wide range of substrates including many xenobiotics, steroids, fatty acids, vitamins and prostaglandins. Depending on the procedure of induction and chromatography numerous forms of enzyme can be isolated. Up to now there is no clear evidence whether a special form of P-450 exists for each of the different reaction types, or several nonspecific enzymes catalyze more than 1 reaction. The enzymes from different organisms and from different tissues of a single organism are not comparable, those described in, #1# &lt;1-3&gt; display only a small part of a wide spectrum of possible reactions) &lt;1-3&gt;</p> <p>P: ?</p>
<b>Source tissue</b>	Liver #1# <1-3>
<b>Localisation</b>	Microsomes #1# (16 different cytochrome P-450 have been isolated from mouse liver of which each contains numerous different forms of P-450) <1, 2>
<b>References</b>	<p>&lt;1&gt; Lang, M.A., Nebert, D.W.: Structural gene products of the Ah locus. Evidence for many unique P-450-mediated monooxygenase activities reconstituted from 3-methylcholanthrene-treated C57BL/6N mouse liver microsomes:: J. Biol. Chem., 256; 12058-12067 (1981)</p> <p>&lt;2&gt; Lang, M.A., Gielen, J.E., Nebert, D.W.: Genetic evidence for many unique liver microsomal P-450-mediated monooxygenase activities in heterogenic stock mice:: J. Biol. Chem., 256; 12068-12075 (1981)</p> <p>&lt;3&gt; Theoharides, A.D., Kupfer, D.: Evidence for different hepatic microsomal monooxygenases catalyzing omega- and (omega-1)-hydroxylations of prostaglandins E1 and E2. Effects of inducers of monooxygenase on the kinetic constants of prostaglandin hydroxylation:: J. Biol. Chem., 256; 2168-2175 (1981)</p>

#### BRENDA:1.14.14.1 Mammals

<b>E.C. number</b>	1.14.14.1 (BRENDA copyright notice)
<b>Original Organism</b>	Mammals (e.g. mouse <1,2>; rabbit <3>; . There is no clear evidence whether the enzymes isolated from microsomes of different species, or even from the same species by different research groups are identical) <1-3>

Substrates/products	<p>2 S: Aryl hydrocarbons + reduced flavoprotein + O<sub>2</sub> (e.g. benzo[a]pyrene, ethoxyresufurin, biphenyl, p-nitroanisole, acetanilide, 2-acetylaminofluorene, 2-ethoxycoumarin, estradiol-17beta, testosterone &lt;1,2&gt;; prostaglandins &lt;3&gt;) &lt;1-3&gt;</p> <p>P: ?</p> <p>2 S: More (acts on a wide range of substrates including many xenobiotics, steroids, fatty acids, vitamins and prostaglandins. Depending on the procedure of induction and chromatography numerous forms of enzyme can be isolated. Up to now there is no clear evidence whether a special form of P-450 exists for each of the different reaction types, or several nonspecific enzymes catalyze more than 1 reaction. The enzymes from different organisms and from different tissues of a single organism are not comparable, those described in &lt;1,2,3&gt;; display only a small part of a wide spectrum of possible reactions) &lt;1-3&gt;</p> <p>P: ?</p>
Source tissue	Liver <1-3>
Localisation	Microsomes (16 different cytochrome P-450 have been isolated from mouse liver of which each contains numerous different forms of P-450) <1, 2>
References	<p>&lt;1&gt; Lang, M.A., Nebert, D.W.: Structural gene products of the Ah locus. Evidence for many unique P-450-mediated monooxygenase activities reconstituted from 3-methylcholanthrene-treated C57BL/6N mouse liver microsomes:: J. Biol. Chem., 256; 12058-12067 (1981)</p> <p>&lt;2&gt; Lang, M.A., Gielen, J.E., Nebert, D.W.: Genetic evidence for many unique liver microsomal P-450-mediated monooxygenase activities in heterogeneic stock mice:: J. Biol. Chem., 256; 12068-12075 (1981)</p> <p>&lt;3&gt; Theoharides, A.D., Kupfer, D.: Evidence for different hepatic microsomal monooxygenases catalyzing omega- and (omega-1)-hydroxylations of prostaglandins E1 and E2. Effects of inducers of monooxygenase on the kinetic constants of prostaglandin hydroxylation:: J. Biol. Chem., 256; 2168-2175 (1981)</p>

<b>Systematic name</b>	Substrate, reduced-flavoprotein:oxygen oxidoreductase (RH-hydroxylating or-epoxidizing)
<b>Recommended name</b>	Unspecific monooxygenase
<b>Synonyms</b>	<ul style="list-style-type: none"> <li><input checked="" type="checkbox"/> Microsomal monooxygenase</li> <li><input checked="" type="checkbox"/> EC 1.99.1.1 (formerly)</li> <li><input checked="" type="checkbox"/> EC 1.14.99.8 (formerly)</li> <li><input checked="" type="checkbox"/> EC 1.14.1.1 (formerly)</li> <li><input checked="" type="checkbox"/> Microsomal P-450</li> <li><input checked="" type="checkbox"/> Aryl hydrocarbon hydroxylase</li> <li><input checked="" type="checkbox"/> Aryl-4-monooxygenase</li> <li><input checked="" type="checkbox"/> Xenobiotic monooxygenase</li> <li><input checked="" type="checkbox"/> Flavoprotein-linked monooxygenase</li> <li><input checked="" type="checkbox"/> Flavoprotein monooxygenase</li> <li><input checked="" type="checkbox"/> Oxygenase, flavoprotein-linked mono-</li> <li><input checked="" type="checkbox"/> EC 1.14.14.2 (formerly)</li> </ul>
<b>CAS registration number</b>	62213-32-5
<b>Reaction</b>	$\text{RH} + \text{reduced flavoprotein} + \text{O}_2 = \text{ROH} + \text{oxidized flavoprotein} + \text{H}_2\text{O}$
<b>Reaction type</b>	<ul style="list-style-type: none"> <li><input checked="" type="checkbox"/> Deamination</li> <li><input checked="" type="checkbox"/> Desulfation</li> <li><input checked="" type="checkbox"/> Epoxidation</li> <li><input checked="" type="checkbox"/> Hydroxylation</li> <li><input checked="" type="checkbox"/> N-, S-, O-Dealkylation</li> <li><input checked="" type="checkbox"/> N-Oxidation</li> <li><input checked="" type="checkbox"/> Redox reaction</li> <li><input checked="" type="checkbox"/> Reduction of azo, nitro, N-oxide groups</li> <li><input checked="" type="checkbox"/> Sulfoxidation</li> </ul>

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## NiceZyme View of ENZYME: EC 1.14.14.1

### Official Name

Unspecific monooxygenase.

### Alternative Name(s)

Microsomal monooxygenase.

Xenobiotic monooxygenase.

Aryl-4-monooxygenase.

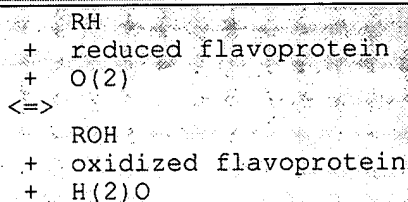
Aryl hydrocarbon hydroxylase.

Microsomal p450.

Flavoprotein-linked monooxygenase.

Cytochrome p450.

### Reaction catalysed



### Cofactor(s)

Heme-thiolate.

### Comments

- Acts on a wide range of substrates including many xenobiotics, steroids, fatty acids, vitamins and prostaglandins.
- Reactions catalysed include hydroxylation, epoxidation, N-oxidation, sulfoxidation, N-, S- and O-dealkylations, desulfation, deamination, and reduction of azo, nitro, and N-oxide groups.

### Cross-References

Biochemical Pathways;  
map number(s)

[T6](#), [U6](#)

PROSITE

[PDOC00081](#)

BRENDA

[1.14.14.1](#)

EMP/PUMA

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WIT	1.14.14.1
KYOTO UNIVERSITY LIGAND CHEMICAL DATABASE	1.14.14.1
MEDLINE	Find literature relating to 1.14.14.1
SWISS-PROT	<p> P24903, C2F1_HUMAN; O18809, C2F3_CAPHI; O35293, C2F4_RAT ;  P33269, C4D1_DROME; O16805, C4D1_DROSI; Q27589, C4D2_DROME;  O18596, C4DA_DROMT; Q27606, C4E2_DROME; P13527, C6A1_MUSDO;  P33270, C6A2_DROME; Q27593, C6A8_DROME; Q27594, C6A9_DROME;  Q04552, C6B1_PAPPO; Q27664, C6B2_HELAM; Q27756, C6B3_PAPPO;  Q27902, C6B4_PAPGL; Q95036, C6B5_PAPGL; Q95031, C6B6_HELAM;  O61387, C6B7_HELAM; P56590, CP11_CANFA; Q06367, CP11_CAVPO;  Q92039, CP11_CHACA; P79716, CP11_DICLA; P04798, CP11_HUMAN;  O42430, CP11_LIMLI; O42231, CP11_LIZAU; P33616, CP11_MACFA;  Q00557, CP11_MESAU; Q92148, CP11_MICTO; P00184, CP11_MOUSE;  Q92110, CP11_ONCMY; Q92095, CP11_OPSTA; P98181, CP11_PAGMA;  Q9YH64, CP11_PLAFE; Q92100, CP11_PLEPL; P05176, CP11_RABIT;  P00185, CP11_RAT ; P56591, CP11_SHEEP; O42457, CP11_SPAAU;  Q92116, CP11_STECH; P56592, CP12_CANFA; Q64391, CP12_CAVPO;  Q01741, CP12_CHICK; P05177, CP12_HUMAN; P24453, CP12_MESAU;  P00186, CP12_MOUSE; P00187, CP12_RABIT; P04799, CP12_RAT ;  Q92109, CP13_ONCMY; P79760, CP14_CHICK; P79761, CP15_CHICK;  Q16678, CP1B_HUMAN; Q64429, CP1B_MOUSE; Q64678, CP1B_RAT ;  P04800, CP31_RAT ; P05183, CP32_RAT ; P05184, CP33_HUMAN;  P08684, CP34_HUMAN; P20815, CP35_HUMAN; P80056, CP35_PAPSP;  P11707, CP36_RABIT; P24462, CP37_HUMAN; P33268, CP38_MACFA;  P51538, CP39_RAT ; Q64148, CP3A_MESAU; Q64459, CP3B_MOUSE;  P24463, CP3C_CANFA; Q64464, CP3D_MOUSE; Q64417, CP3E_CAVPO;  Q64406, CP3F_CAVPO; Q64481, CP3G_MOUSE; Q64409, CP3H_CAVPO;  Q64581, CP3I_RAT ; P79152, CP3J_CAPAE; O18993, CP3L_CALJA;  Q29496, CP3O_SHEEP; O09158, CP3P_MOUSE; O42563, CP3R_ONCMY;  P79102, CP3S_BOVIN; P79401, CP3T_PIG ; O70537, CP3V_MESAU;  P10611, CP44_RABIT; P24464, CP48_RAT ; P13584, CP4B_HUMAN;  Q64462, CP4B_MOUSE; P15128, CP4B_RABIT; P15129, CP4B_RAT ;  P29981, CP4C_BLADI; P10613, CP51_CANAL; P50859, CP51_CANGA;  P14263, CP51_CANTR; Q16850, CP51_HUMAN; Q02315, CP51_ISSOR;  Q12664, CP51_PENIT; O46420, CP51_PIG ; Q64654, CP51_RAT ;  Q09736, CP51_SCHPO; P93846, CP51_SORBI; O14442, CP51_UNCNE;  P49602, CP51_USTMA; P93596, CP51_WHEAT; P10614, CP51_YEAST;  P21595, CP56_YEAST; P10615, CP5A_CANTR; P30607, CP5B_CANTR;  P16496, CP5C_CANMA; P16141, CP5D_CANMA; P24458, CP5E_CANMA;  P30608, CP5F_CANTR; P30609, CP5G_CANTR; P30610, CP5H_CANTR;  Q12586, CP5I_CANMA; Q12588, CP5J_CANMA; Q12589, CP5K_CANMA;  P30611, CP5N_CANTR; P43083, CP5V_CANAP; Q12573, CP5W_CANAP;  Q05047, CP72_CATRO; P11711, CPA1_RAT ; P15149, CPA2_RAT ;  P20812, CPA3_RAT ; P15392, CPA4_MOUSE; P20852, CPA5_MOUSE;  P11509, CPA6_HUMAN; P20853, CPA7_HUMAN; P80055, CPA7_PAPSP;  P24454, CPA8_MESAU; P24455, CPA9_MESAU; Q05555, CPAA_RABIT;  Q05556, CPAB_RABIT; P56593, CPAC_MOUSE; Q16696, CPAD_HUMAN;  P22779, CPAX_BOVIN; P00176, CPB1_RAT ; P04167, CPB2_RAT ;  P13107, CPB3_RAT ; P00178, CPB4_RABIT; P12789, CPB5_RABIT;  P20813, CPB6_HUMAN; P12790, CPB9_MOUSE; P12791, CPBA_MOUSE;  P24460, CPBB_CANFA; P33272, CPBC_RAT ; O55071, CPBJ_MOUSE;  Q62397, CPBK_MOUSE; P34033, CPBX_CAVPO; P00180, CPC1_RABIT;  P00181, CPC2_RABIT; P00182, CPC3_RABIT; P11371, CPC4_RABIT;  P00179, CPC5_RABIT; P05178, CPC6_RAT ; P05179, CPC7_RAT ;  P10632, CPC8_HUMAN; P11712, CPC9_HUMAN; P11713, CPCA_HUMAN;  P08683, CPCL_RAT ; P11510, CPCC_RAT ; P20814, CPCD_RAT ;  P17666, CPCE_RABIT; P11372, CPCF_RABIT; P15123, CPCG_RABIT;  P33260, CPCI_HUMAN; P33261, CPCJ_HUMAN; P33262, CPCK_MACFA;  P56594, CPCL_CANFA; P19225, CPCM_RAT ; P24470, CPCN_RAT ;  P33273, CPCO_RAT ; Q08078, CPCP_MESAU; P33263, CPCQ_MESAU; </p>

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P10634, CPD2_RAT ;	P12938, CPD3_RAT ;	P13108, CPD4_RAT ;
P12939, CPD5_RAT ;	P10635, CPD6_HUMAN;	P11714, CPD9_MOUSE;
P24456, CPDA_MOUSE;	P24457, CPDB_MOUSE;	Q01361, CPDE_BOVIN;
Q29473, CPDF_CANFA;	Q64403, CPDG_CAVPO;	Q29488, CPDH_MACFA;
Q64680, CPDI_RAT ;	Q18992, CPDJ_CALJA;	Q18963, CPE1_BOVIN;
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Q05421, CPE1_MOUSE;	P79383, CPE1_PIG ;	P08682, CPE1_RABIT;
P05182, CPE1_RAT ;	P33274, CPF1_RAT ;	P51869, CPF4_RAT ;
P51870, CPF5_RAT ;	P51871, CPF6_RAT ;	P24461, CPG1_RABIT;
P10610, CPG1_RAT ;	P05180, CPH1_CHICK;	P20678, CPH2_CHICK;
P52786, CPJ1_RABIT;	P51589, CPJ2_HUMAN;	P51590, CPJ3_RAT ;
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Q93299, CPK3_ONCMY;	Q93297, CPK4_ONCMY;	Q27712, CPL1_PANAR;
Q92088, CPM1_ONCMY;	P46194, CPV1_BOVIN;	Q42145, CPV1_BRARE;
P79690, CPV1_CARAU;	P19098, CPV1_CHICK;	P79699, CPV1_COTJA;
Q46512, CPV1_HORSE;	P11511, CPV1_HUMAN;	Q92111, CPV1_ICTPU;
P28649, CPV1_MOUSE;	P70091, CPV1_ORENI;	Q92087, CPV1_ORYLA;
Q29624, CPV1_PIG ;	Q92112, CPV1_POEGU;	Q29605, CPV1_RABIT;
P22443, CPV1_RAT ;	Q9XS28, CPV1_SHEEP;	Q73686, CPV2_CARAU;
P79430, CPV2_PIG ;	P79304, CPV3_PIG ;	P14779, CPXB_BACME;
P14762, CPXI_BACME;	P56654, CPZ2_MOUSE;	P56655, CPZ3_MOUSE;
P56656, CPZ4_MOUSE;	P56657, CPZ5_MOUSE;	Q62671, CPZ6_CANFA;
P79402, CPZ7_PIG ;		

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## BRENDA:1.3.3.6

E.C. number	1.3.3.6 (BRENDA <a href="#">copyright notice</a> )
Original Organism	<p><b>#1#</b> <a href="#">Spinacia oleracea</a> (spinach) &lt;20&gt;</p> <p><b>#2#</b> <a href="#">Vigna radiata</a> (mung bean) &lt;20&gt;</p> <p><b>#3#</b> <a href="#">Phanerochaete chrysosporium</a> &lt;21&gt;</p> <p><b>#4#</b> <a href="#">Cucumis sativus</a> (cucumber) &lt;22&gt;</p> <p><b>#5#</b> <a href="#">Candida tropicalis</a> (pK 233) &lt;1, 13-16&gt;</p> <p><b>#6#</b> Rat (3 forms: 1. inducible fatty acyl-CoA oxidase, 2. noninducible fatty acyl-CoA oxidase, 3. noninducible trihydroxycoprostanoyl-CoA oxidase &lt;11&gt;) &lt;2-4, 6-11&gt;</p> <p><b>#7#</b> <a href="#">Candida lipolytica</a> &lt;5&gt;</p> <p><b>#8#</b> <a href="#">Candida maltosa</a> &lt;18&gt;</p> <p><b>#9#</b> <a href="#">Human</a> &lt;12&gt;</p> <p><b>#10#</b> <a href="#">Saccharomyces cerevisiae</a> &lt;17&gt;</p> <p><b>#11#</b> <a href="#">Mouse</a> &lt;19&gt;</p>
Systematic name	Acyl-CoA:oxygen 2-oxidoreductase
Recommended name	Acyl-CoA oxidase
Synonyms	<p> Oxidase, acyl-coenzyme A</p> <p> Fatty acyl-CoA oxidase</p> <p> Acyl coenzyme A oxidase</p> <p> Fatty acyl-coenzyme A oxidase</p>
CAS registration number	61116-22-1
Reaction	Acyl-CoA + O <sub>2</sub> = trans-2,3-dehydroacyl-CoA + H <sub>2</sub> O <sub>2</sub> (acts on CoA derivatives of fatty acids with chain length from 8 to 18)
Reaction type	<p> More <b>#7#</b> (anti-elimination of pro-2R and pro-3R hydrogens of acyl-CoA) &lt;5&gt;</p> <p> Redox reaction</p>
Substrates/products	<p> S: Lauroyl-CoA + O<sub>2</sub> <b>#1,2,5,6#</b> &lt;6, 13, 15, 20&gt; P: trans-2-Dodecenoyl-CoA + H<sub>2</sub>O<sub>2</sub></p> <p> S: Arachidoyl-CoA + O<sub>2</sub> <b>#5#</b> &lt;13&gt; P: trans-2,3-Dehydro-5,8,11,14-eicosatetraenoyl-CoA + H<sub>2</sub>O<sub>2</sub></p> <p> S: Dec-4-cis-enoyl-CoA + O<sub>2</sub> <b>#6#</b> &lt;3&gt; P: ?</p> <p> S: 2-Oxoheptadecyldethio-CoA + O<sub>2</sub> <b>#5#</b> &lt;14&gt;</p>

P: ?

Ⓢ S: Dicarboxylic acid-CoAs with 6-16 carbon atoms + O<sub>2</sub> #6# <10>

P: ?

Ⓢ S: Butyryl-CoA + O<sub>2</sub> #1,2# (not, #3,6# <9, 21>) <20>

P: trans-2-Butenoyl-CoA + H<sub>2</sub>O<sub>2</sub>

Ⓢ S: Linoleoyl-CoA + O<sub>2</sub> #1,2,6# <6, 20>

P: trans-2,3-Dehydro-9,12-octadienoyl-CoA + H<sub>2</sub>O<sub>2</sub>

Ⓢ S: Oleoyl-CoA + O<sub>2</sub> #5,6# <6, 13>

P: trans-2,3-Dehydro-9-octadecenoyl-CoA + H<sub>2</sub>O<sub>2</sub>

Ⓢ S: Myristoyl-CoA + O<sub>2</sub> #1,2,5,6# <6, 13, 20>

P: trans-2-Tetradecenoyl-CoA + H<sub>2</sub>O<sub>2</sub>

Ⓢ S: Octanoyl-CoA + O<sub>2</sub> #1,2,5,6# <6, 13, 20>

P: trans-2-Octenoyl-CoA + H<sub>2</sub>O<sub>2</sub>

Ⓢ S: Leuko-dichlorofluorescein + O<sub>2</sub> #6# <7>

P: ?

Ⓢ S: Nonanoyl-CoA + O<sub>2</sub> #7# <5>

P: trans-2-Nonenoyl-CoA + H<sub>2</sub>O<sub>2</sub>

Ⓢ S: Stearoyl-CoA + O<sub>2</sub> #1,5-7# <5, 6, 13, 20>

P: trans-2-Octadecenoyl-CoA + H<sub>2</sub>O<sub>2</sub>

Ⓢ S: Hexadecanedioyl-CoA + O<sub>2</sub> #6# <4>

P: ?

Ⓢ S: Palmitoyl-CoA + O<sub>2</sub> #1,2,5,6# <4, 6, 7, 9, 13, 20>

P: trans-2,3-Dehydrohexadecanoyl-CoA + H<sub>2</sub>O<sub>2</sub>  
(trans-2-hexadecenoyl-CoA)

Ⓢ S: Acyl-CoA + O<sub>2</sub> #1-7# (specificity: C4-C20 acyl-CoA, #5# <13>; C8-C18 acyl CoA, #3# <21>; most active towards C12-C18 acyl-CoA, C20 and C22 acyl-CoA also oxidized, C4 and C6 acyl-CoA hardly oxidized, #6# <9>; C4-C18 monocarboxylic acid-CoA, #6# <10>; C6-C16 dicarboxylic-CoA, #6# <10>; 3'-phosphate on the ribose ring and the structure of the adenine moiety are essential for substrate recognition, specificity is relatively low with respect to the structure of the pantonic acid moiety, #6# <6>; chain-length specificity changes with acyl-CoA concentration used, #6# <7>; Cucumis sativus: enzyme acts selectively on fatty acyl-CoA with 16 or 18 carbon atoms, cis-9-unsaturated esters with a C16 or C18 acyl moiety being converted with higher rate than saturated or polyunsaturated fatty acyl-CoA, #4# <22>; anti-elimination of pro-2R and pro-3R hydrogens of acyl-CoA, #7# <5>) <3-9, 10, 13-15, 20-22>

	<p>P: trans-2,3-Dehydroacyl-CoA + H<sub>2</sub>O<sub>2</sub></p> <p>☒ S: Trihydroxycoprostanoyl-CoA + O<sub>2</sub> #6# &lt;4&gt;</p> <p>P: ?</p> <p>☒ S: Decanoyl-CoA + O<sub>2</sub> #5,6# &lt;6, 13&gt;</p> <p>P: trans-2-Decenoyl-CoA + H<sub>2</sub>O<sub>2</sub></p>
Natural substrates	<p>Acyl-CoA + O<sub>2</sub> #5,6,9# (CoA derivatives of fatty acids with chain length from 8 to 18, first reaction of peroxisomal beta-oxidation, rate limiting for this process, #6,9# &lt;7, 8, 12&gt;; beta-oxidation of dicarboxylic acid-CoAs in rat liver is carried out exclusively in peroxisomes, #6# &lt;10&gt;; significance in metabolism of alkanes of <i>Candida tropicalis</i>, #5# &lt;16&gt;) &lt;7, 8, 10, 12, 16&gt;</p>
Turnover number (1/min)	<p>-999 #5# (pH-dependency of turnover number) &lt;15&gt;</p>
Specific activity (micromol/min/mg)	<p>☒ 27.2 #6# &lt;8&gt;</p> <p>☒ 19.13 #5# &lt;13&gt;</p> <p>☒ 2.04 #6# &lt;6&gt;</p> <p>☒ 1.45 #6# &lt;9&gt;</p> <p>☒ -999 #5# &lt;14&gt;</p>
K <sub>m</sub> -value (mM)	<p>☒ 0.087 #6# {octanoyl-CoA} (liver enzyme) &lt;7&gt;</p> <p>☒ 0.058 #6# {octanoyl-CoA} &lt;6&gt;</p> <p>☒ 0.046 #5# {oleoyl-CoA} &lt;13&gt;</p> <p>☒ 0.042 #5# {octanoyl-CoA} &lt;13&gt;</p> <p>☒ 0.034 #5# {stearoyl-CoA} &lt;13&gt;</p> <p>☒ 0.0335 #5# {arachidoyl-CoA} &lt;13&gt;</p> <p>☒ 0.032 #1# {butyryl-CoA} &lt;20&gt;</p> <p>☒ 0.029 #5# {myristoyl-CoA} &lt;13&gt;</p> <p>☒ 0.027 #6# {lauroyl-CoA} (liver enzyme) &lt;6&gt;</p> <p>☒ 0.025 #5# {dodecanoyl-CoA} (pH 7-9.5) &lt;15&gt;</p> <p>☒ 0.024 #5# {decanoyl-CoA} (lauroyl-CoA) &lt;13&gt;</p> <p>☒ 0.023 #1# {stearoyl-CoA} &lt;20&gt;</p> <p>☒ 0.02 #6# {decanoyl-CoA} &lt;6&gt;</p> <p>☒ 0.019 #1# {linoleoyl-CoA} &lt;20&gt;</p> <p>☒ 0.013 #6# {lauroyl-CoA} &lt;6&gt;</p> <p>☒ 0.0116 #6# {myristoyl-CoA} (palmitoyl-CoA) &lt;6&gt;</p> <p>☒ 0.011 #1,6# {myristoyl-CoA} (#1# &lt;20&gt;; oleoyl CoA, #6# &lt;6&gt;) &lt;6, 20&gt;</p> <p>☒ 0.0096 #6# {stearoyl-CoA} &lt;6&gt;</p>

	<ul style="list-style-type: none"> <li>■ 0.0093 #6# {dec-4-cis-enoyl-CoA} &lt;3&gt;</li> <li>⊗ 0.0073 #6# {linoleoyl-CoA} &lt;6&gt;</li> <li>⊗ 0.007 #6# {palmitoyl-CoA} (liver enzyme) &lt;7&gt;</li> <li>⊗ 0.005 #6# {O2} &lt;6&gt;</li> <li>⊗ 0.00181 #6# {palmitoyl-CoA} (kidney enzyme) &lt;7&gt;</li> </ul>
<b>pH-optimum</b>	8 #5,6# <6, 13>
<b>pH-range</b>	7-10 #6# (7: about 30% of activity maximum, 10: about 5% of activity maximum, inactive below pH 6.5) <6>
<b>Temperature-optimum (deg.C)</b>	50 #5# <13>
<b>Cofactors/prosthetic groups</b>	FAD #5,6# (flavoprotein, #5,6# <6, 7, 13-15>; prosthetic group, #6# <6>; 1.22 mol per mol of enzyme, #6# <6>; flavoprotein with noncovalently bound FAD, #6# <7>; 8 mol FAD per mol of enzyme, 1 mol FAD per mol of subunit, #5# <13, 14>) <6, 7, 13-15>
<b>Inhibitors</b>	<ul style="list-style-type: none"> <li>⊗ 3-Ketohexadecanoyl-CoA #6# &lt;6&gt;</li> <li>⊗ Acetyl-CoA #6# &lt;3&gt;</li> <li>⊗ AgNO3 #5# &lt;13&gt;</li> <li>⊗ Antimycin A #6# &lt;4&gt;</li> <li>⊗ C16-C18 fatty acyl-CoA #11# (at fairly low concentrations) &lt;19&gt;</li> <li>⊗ CH3COOK #6# &lt;6&gt;</li> <li>⊗ CoA #6# &lt;3&gt;</li> <li>⊗ FMN #6# &lt;3&gt;</li> <li>⊗ HgCl2 #5# &lt;13&gt;</li> <li>⊗ KBr #6# &lt;6&gt;</li> <li>⊗ KCl #6# &lt;6&gt;</li> <li>⊗ KI #6# &lt;6&gt;</li> <li>⊗ KNO3 #6# &lt;6&gt;</li> <li>⊗ Mercuric acetate #5# &lt;13&gt;</li> <li>⊗ NaCl #6# &lt;6&gt;</li> <li>⊗ NaN3 #6# &lt;6&gt;</li> <li>⊗ NH4Cl #6# &lt;6&gt;</li> <li>⊗ NH4SCN #6# &lt;6&gt;</li> <li>⊗ p-Chloromercuribenzoate #5# &lt;13&gt;</li> </ul>

<b>Source tissue</b>	<input checked="" type="checkbox"/> Adrenal gland #6# <7> <input checked="" type="checkbox"/> Heart #6# <7> <input checked="" type="checkbox"/> Kidney #6,11# <7, 19> <input checked="" type="checkbox"/> Liver #6,9# <3, 6-12> <input checked="" type="checkbox"/> Mycelium #3# <21> <input checked="" type="checkbox"/> Seedlings #4# (cotyledons) <22> <input checked="" type="checkbox"/> Skeletal muscle #6# <7>
<b>Localisation</b>	<input checked="" type="checkbox"/> Extracellular #3# <21> <input checked="" type="checkbox"/> Glyoxysomes #4# <22> <input checked="" type="checkbox"/> Peroxisomes #1,2,5,6,9,10# <4, 6, 8, 10-12, 15-17, 20>
<b>Purification</b>	<input checked="" type="checkbox"/> #4# (cucumber) <22> <input checked="" type="checkbox"/> #5# <13, 15> <input checked="" type="checkbox"/> #6# <6, 8, 9>
<b>Crystallisation</b>	#5# <13>
<b>Molecular Weight</b>	<input checked="" type="checkbox"/> 600000 #5# (Candida tropicalis, sedimentation equilibrium) <13> <input checked="" type="checkbox"/> 552000 #5# (Candida tropicalis, ultracentrifugation) <15> <input checked="" type="checkbox"/> 427000 #6# (rat liver, noninducible fatty acyl-CoA oxidase, gel filtration) <11> <input checked="" type="checkbox"/> 150000 #4# (Cucumis sativus, gel filtration) <22> <input checked="" type="checkbox"/> 145000 #6# (rat liver, inducible fatty acyl-CoA oxidase, gel filtration) <11> <input checked="" type="checkbox"/> 139000 #6# (rat liver, sedimentation equilibrium method <6>; noninducible trihydroxycoprostanoyl-CoA oxidase <11>) <6, 11>
<b>Subunits</b>	<input checked="" type="checkbox"/> Dimer #4,6# (2 * 71000, rat liver, SDS-PAGE, noninducible fatty acyl-CoA oxidase <11>; 2 * 72000, Cucumis sativus, SDS-PAGE <22>) <11, 22> <input checked="" type="checkbox"/> Hexamer #6# (6 * 69000, rat liver, SDS-PAGE, noninducible trihydroxyprostanoyl-CoA oxidase) <11> <input checked="" type="checkbox"/> Octamer #5# (8 * 74000 <13>; 8 * 75000 <14>; Candida tropicalis, SDS-PAGE <13, 14>) <13, 14> <input checked="" type="checkbox"/> Oligomer #5,6# (x * 72100, Candida tropicalis, SDS-PAGE <15>; x * 52000, x * 22500, rat liver, SDS-PAGE, inducible fatty acyl-CoA oxidase <11>) <11, 15> <input checked="" type="checkbox"/> Tetramer #6# (2 * 45000 + 2 * 22000, rat, SDS-PAGE) <8>
<b>Cloned</b>	#5,6# (Candida tropicalis pK 233 <1>; rat <2>) <1, 2>
<b>pH-stability</b>	5.5-9 #5# (35°C, 60 min) <13>

<b>Temperature stability (deg.C)</b>	<p>☑ 65 #5# (10 min, complete inactivation) &lt;13&gt;</p> <p>☑ 50 #5# (10 min) &lt;13&gt;</p>
<b>General stability</b>	#4# Dialysis, 4°C, 24 h, 80% loss of activity <22>
<b>Storage stability</b>	<p>☑ #5# -20°C, 25% glycerol, 8 months &lt;15&gt;</p> <p>☑ #6# -20°C, for at least 1 month &lt;6&gt;</p> <p>☑ #5# -20°C, pH 7.4, 2 months &lt;14&gt;</p> <p>☑ #5# 4°C, pH 7.4, 2 months, 20% loss of activity &lt;14&gt;</p> <p>☑ #4# Frozen, 10% sucrose, several weeks &lt;22&gt;</p>
<b>References</b>	<p>&lt;1&gt; Rachubinski, R.A., Fujiki, Y., Lazarow, P.B.: Cloning of cDNA coding for peroxisomal acyl-CoA oxidase from the yeast <i>Candida tropicalis</i> pK233:: Proc. Natl. Acad. Sci. USA, 82; 3973-3977 (1985)</p> <p>&lt;2&gt; Osumi, T., Ozasa, H., Hashimoto, T.: Molecular cloning of cDNA for rat acyl-CoA oxidase:: J. Biol. Chem., 259; 2031-2034 (1984)</p> <p>&lt;3&gt; Hovik, R., Osmundsen, H.: A kinetic investigation of the acyl-CoA oxidase reaction with the use of a novel spectrophotometric assay. Inhibition by acetyl-CoA, CoA and FMN:: Biochem. J., 263; 297-299 (1989)</p> <p>&lt;4&gt; Vamecq, J., Schepers, L., Parmentier, G., Mannaerts, G.P.: Inhibition of peroxisomal fatty acyl-CoA oxidase by antimycin A:: Biochem. J., 248; 603-607 (1987)</p> <p>&lt;5&gt; Kawaguchi, A., Tsubotani, S., Seyama, Y., Yamakawa, T., Osumi, T., Hashimoto, T., Kikuchi, T., Ando, M., Okuda, S.: Stereochemistry of dehydrogenation catalyzed by Acyl-CoA oxidase:: J. Biochem., 88; 1481-1486 (1980)</p> <p>&lt;6&gt; Osumi, T., Hashimoto, T., Ui, N.: Purification and properties of acyl-CoA oxidase from rat liver:: J. Biochem., 87; 1735-1746 (1980)</p> <p>&lt;7&gt; Reubsæet, F.A.G., Veerkamp, J.H., Bukkens, S.G.F., Trijbels, J.M.F., Monnens, L.A.H.: Acyl-CoA oxidase activity and peroxisomal fatty acid oxidation in rat tissues:: Biochim. Biophys. Acta, 958; 434-442 (1988)</p> <p>&lt;8&gt; Inestrosa, N.C., Bronfman, M., Leighton, F.: Purification of the peroxisomal fatty acyl-CoA oxidase from rat liver:: Biochem. Biophys. Res. Commun., 95; 7-12 (1980)</p> <p>&lt;9&gt; Osumi, T., Hashimoto, T.: Acyl-CoA oxidase of rat liver: a new enzyme for fatty acid oxidation:: Biochem. Biophys. Res. Commun., 83; 479-485 (1978)</p> <p>&lt;10&gt; Suzuki, H., Yamada, J., Watanabe, T., Suga, T.:</p>

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